Expression Patterns of Galectins-1, -3, and -7 Are are Prognostic Markers for Overall Survival of in Ovarian Cancer Patients

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Please be sure to provide author names, affiliations, and contact information according to the journal's guidelines (the corresponding author should be identified with an asterisk).

Finally, please make sure you go through the Scientific Editing Report and incorporate the recommendations.

Abstract: There is a tremendous considerable need for the development ofing new useful prognostic factors in ovarian cancer. Galectins are a family of carbohydrate--binding proteins which that have been suggested to serve as prognostic factors for various cancer types. In this study, the presence expression of gGalectin (Gal)-1, -3, and -7 was investigated in 156 ovarian cancer specimens by using immunohistochemical staining. Staining was evaluated in the cytoplasm and nucleus of cancer cells as well as the peritumoral stroma using a semi quantitative score (Remmele (IR) score). Patients' oOverall patient survival was compared between among different groups of stratified by G galectin expression. Galectin (Gal)-1 and -3 staining was observed in the peritumoural stroma as well as the nucleus and cytoplasm of tumour cells, while Gal-7 was only present in the cytoplasm-of tumor cells. Patients with Gal-1 expression in the cytoplasm or high Gal-1 expression in the peritumoural stroma showed reduced overall survival. Nuclear Gal-3 staining correlated with a better clinical outcomes. We observed a significantly reduced overall survival for c Cases with high Gal-7 expression exhibited significantly reduced overall survival, while and a better survival for-Gal-7negative cases exhibited improved survival, when compared to cases with low expression of Gal-7. We were able to show that bOur results indicate that oth tumour and stromal staining of Gal-1 and cytoplasmic staining of Gal-7 could-serve as negative prognostic factorss for ovarian cancer, while nuclear . We were able to confirm cytoplasmic Gal-7 as a negative prognostic factor. Gal-3 staining in the nucleus could may represent be a new positive prognosticator for ovarian cancer. These findings suggest that galectins may represent promising new targets for ovarian cancer treatment.

Keywords:

Galectin-1; Galectin-3; Galectin-7; ovarian cancer; overall survival

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Recommended actions:

While background information on galectins is provided, the rationale for conducting this study and its novelty are not explicitly stated in the abstract. Include a statement for this, while framing existing information more concisely to adhere to the journal word limit.

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1. Introduction

Ovarian cancer is the most lethal gynecological malignancy, ranking fifth in estimated cancer deaths among women in the USA [1]. First-line treatment consists of primary debulking surgery followed by platinum and paclitaxel chemotherapy² [2]. StillDespite these treatments, the 5-year relative survival rate for epithelial ovarian cancer patients is remains belowless than 50%³ [2]. A lack of screening methods and the frequent presentation with advanced stage disease are considered as the main reasons for the poor outcomes of ovarian cancer patients.

<u>Prognosticators in ovarian cancer include Dd</u>isease stage at diagnosis, extent of residual disease after surgery, histological subtype, and <u>a highthe</u> volume of ascites⁴-can be used as prognosticators in ovarian cancer [4]. Numerous studies have aimed to introduce-identify new biological prognostic factors in ovarian cancer. Recently, the carbohydrate stem cell marker TF1 has been proposed as <u>a</u> negative prognostic marker in ovarian cancer displaying wild_ type p53, while estrogen receptor promoter methylation <u>could</u>-predicts overall survival in low-grade ovarian carcinoma patients^{5.6} [5,6]. Although for these and various other molecules the prognostic value independently of clinical parameters has been provendemonstrated for these and various other molecules, until todayto date, with the exception of for_breast cancer gene (*BRCA*)_-status, no biological marker is commonly accepted⁴ [4]. Further specification of anticancer therapiesy necessitatesarily requires an improvement of in the biological prognostic markers in for ovarian cancer.

Galectins have been defined as belong to a family of proteins sharing two main characteristics: a-binding affinity for β -galactosides and a-significant similarity in the carbohydrate-recognition domain (CRD)⁷ [7]. The first member of this family to be described was gGalectin (Gal)-1, which is can be isolated as a homodimers composed of comprising -two identical CRD subunits⁸ [8]. Since then, a growing number of the gGalectin family members haves had a growing number of members been identified, but only Galectin (Gal)-1–4, Gal-7– 10, Gal-12, and Gal-13 are known to be present in humans⁹ [9]. Similar to Gal-1, Gal-7 typically occurs in-as a homodimers, while Gal-3 is the only gGalectin characterized as a chimeric protein that is known to form higher order oligomers^{10,11} [10,11]</sup>. In several types of cancer types, gGalectins are known to affect tumour growth, metastasis, angiogenesis, cell migration, as well as tumor invasiveness, and progression, and they are therefore very likelygood candidates for proteins with to show a prognostic value for patients² survival^{9,12} [9,12].

The role of <u>GalectinGal</u>-1 in cancer has been studied by various groups, <u>and several</u> papers already exist on this topic. For <u>In</u> patients² sera and ovarian cancer tissues, it has been shown that a combination of CA-125 and Galectin-1 serves as a possible two-marker combination for <u>the</u> preoperative discrimination of benign and malignant ovarian masses [<u>131</u>¹³. <u>AlsoIn addition</u>, patients suffering from metastatic epithelial ovarian cancer were observed to <u>show exhibit</u> higher serum Gal-1 levels than those with non-metastatic typecancer. Elevated <u>Gal-1 staining of the</u> peritumoural stroma staining of Gal-1 was shown

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Recommended actions:

Information that is not relevant to understanding the rationale for the study, such as information on Gal oligomerization or the number of CRD domains, should be removed to make this section succinct.

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to occur in advanced stages of epithelial ovarian cancer and is also <u>connected associated</u> with <u>poorer-reduced</u> progression-free survival in univariate analysis¹⁴ [14]. However, these results have not yet been reproduced for overall survival or confirmed by multivariate analysis¹⁵ [15]. <u>Due to this Thus</u>, the <u>possibility potential</u> of Gal-1 as an independent prognostic marker in ovarian cancer still needs to be requires further investigationed.

High cytoplasmic Galectin-3 expression has been suggested as a negative prognostic factor, as it was shown to correlate with <u>shorter-reduced</u> progression-free survival in ovarian cancer¹⁶ [16]. However, in another study, Gal-3 expression did not correlate to<u>with</u> reduced overall survival, <u>but though</u> a cytoplasmic staining pattern was associated with poor outcome when compared to patterns including nuclear staining¹⁷ [17]. Although Gal-3 staining <u>has</u> <u>been observed</u> in <u>the</u> nucleus and stroma-has been observed, <u>their its</u> influence on overall survival <u>still maintainsremains</u> elusiveunclear.

Finally, Galectin-7 has been proposed by two independent groups to serve as a negative prognostic factor in ovarian cancer by two independent groups. In both studies, its influence on progression-free survival and overall survival has beenwas confirmed by univariate and multivariate analysis^{16.18} [16,18]. YetHowever, there is further disagreement remains regarding whether Gal-7 staining occurs predominantly in the nucleus or the cytoplasm. In additionAlso, it remains is currently unknown if whether there is a correlation between the expressions of different gGalectins are correlated in ovarian cancer, and there is a critical desperate-need for a comprehensive studyies of various gGalectins on in a representative ovarian cancer panel. Therefore, in this study, we investigated the prognostic influence-value of Gal-1, -3, and -7 in patients with epithelial ovarian cancer and analyszed correlations to each otheramong the expression patterns of the three proteins and as well as to-with clinical and pathological parameters. We hypothesizeOur results suggest that Gal-1, -3, and -7 are localization-dependent prognostic factors for overall survival in ovarian cancer patients.

2. Results

2.1. Gal-1 <u>t</u>Fumo<u>u</u>r and <u>s</u>Stroma<u>l</u> <u>s</u>Staining <u>i</u>Is <u>a Nn</u>egative <u>Pp</u>rognostic <u>indicator of for</u> <u>Oo</u>verall <u>Ss</u>urvival

Galectin-1 staining was successfully performed onconducted in 150 ovarian cancer specimens. Gal-1 was present in the cytoplasms and the nuclei of ovarian cancer cells, as well as in the peritumoural stromae (Figure 1Fig. 1). In 102 cases (68.0%), the eytoplasms of tumour cell_cytoplasms were-was positive for Gal-1, with a median Remmele immunoreactive (IR) score-(IRS) of 3. The Peperitumoural stroma was positive for Gal-1 in 148 cases (98.0%), with a median IR scores of 8. Gal-1 expression was significantly correlated with several clinical and pathological data-factors (Table 1Table 1).

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Insufficient background information and literature review

Recommended actions:

The Introduction does not provide a sufficient background of the problem studied. The biological functions of galectins related to tumorigenesis, including malignant transformation, invasion, and metastasis, are not described, and it is unclear how galectins are involved in all these processes. I have provided recommendations in this regard along with suggestions for additional papers that can be cited in the *High Impact Peer Review Report*. These points should be discussed in the manuscript.

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Restructuring of information in the Results section

Recommended actions:

The results section requires greater re-organization, because the presentation of the data does not correspond to their placement in your illustrations. I have elaborated on this in my assessment of your manuscript.

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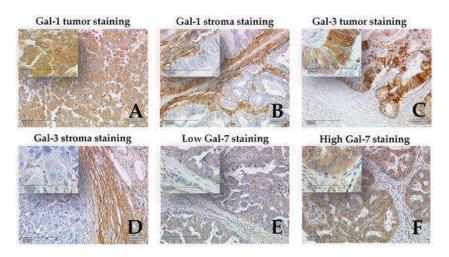


Figure 1. Detection of <u>gGalectins</u> by immunohistochemistry. Representative photomicrographs are shown. <u>Galectin (Gal)-1</u> was present in the cytoplasm and the nucle<u>usi</u> of ovarian cancer cells (**A**) as well as the peritumo<u>u</u>ral stroma (**B**)_{<u>i</u>; Gal-3 staining was observed in the nucle<u>usi</u>, cytoplasm (**C**), and stroma (**D**)_{<u>i</u>; Staining for Galectin-7 was mainly observed in the cytoplasm (**E**), with; only a few individual cases show<u>inged</u> nuclear staining (**F**)_{<u>i</u>; 10× magnification was used for the outer pictures and 50× magnification for the inserts The sScale bars, in in the outer pictures equal-200 µm (10× magnification) in main images, and the scale bars in the inserts equal-100 µm (50× magnification) in insets.}}}

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 Table 1. Correlations between Gal-1 staining correlated with and clinical and pathological data factors.

Clinical and Pathological Variables	Gal-1 Expression Cytoplasm		P	Gal-1 Expression Stroma		p	Gal-1 Expression Nucleus		p
	negative	positive		low	high		negative	positive	
Histology									
Serous	26	79	0.008	34	71	NS	27	78	0.002
Clear cell	5			6	6		3	9	
Endometrioid	8	12		6 7 3	13		11	9 9 4	
Mucinous	9	4		3	10		9	4	
Tumor Stage									
pT1	22	17	< 0.001	20	19	0.006	19	20	0.020
pT2+	26	84		30	80		31	79	
Lymph node									
pN0/pNX	36	65	NS	34	67	NS	43	58	0.001
pN1	12	37		16	33		7	42	
Distant Metastasis									
pM0/pMX	47	97	NS	49	95 5	NS	49	95	NS
pM1	1	5		1	5		1	95 5	
Grading									
GI	20	16	< 0.001	13	23	NS	14	22	NS
G2+	22	80		31	71		31	71	
FIGO									
1/11	22	21	0.001	17	26	NS	21	22	0.013
III/IV	24	78		31	71		28	74	
Age									
≤60 years	27	52	NS	28	51	NS	24	55	NS
<60 years	21	50		22	49	20/330	26	45	

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TNM staging was accomplished performed according to the actual standards of the Union for International Cancer Control (UICC); pT1 = tumour stage 1; pT2+ = tumour stage 2 or higher; pN0 = lymph node stage 0; pNX = lymph node stage not evaluated; pN1 = lymph node stage 1; pM0 = distant metastasis stage 0; pMX = distant metastasis not evaluated; pM1 = distant metastasis stage 1; G1 = grade 1; G2+ = grade 2 or higher; FIGO = Fédération Internationale de Gynécologie et d'Obstétrique; NS = Not significant (p > 0.05)

Gal-1 staining in the cytoplasm and nucleus showed differedness for among several histological subtypes (p = 0.008 and, p = 0.002, respectively). Cytoplasmic Gal-1 staining was significantly stronger in serous, clear cell, or endometrioid subtypes, while for the mucinous subtype, we found observed more negative cases. AlsoIn addition, more cases showed Gal-1 positive nuclei for with serous and clear cell subtypes exhibited Gal-1-positive nuclei, while the endometrioid and mucinous subtypes had exhibited weaker nuclear Gal-1 stainings. Furthermore, Gal-1 staining in the nucleus, cytoplasm, and stroma were was significantly higher in cases with advanced tumour stage (p < 0.001, p = 0.006, and p = 0.02, respectively). Gal-1 expression in the cytoplasm was significantly higher in cases with higher grading (p < 0.001) and advanced FIGO (Fédération Internationale de Gynécologie et d'Obstétrique) stage (p = 0.001). The IR scores of nuclear Gal-1 staining in the nucleus showed were higher IR scores in lymph node_positive cases (p = 0.001) and eases those with advanced FIGO stage (p = 0.013).

<u>The Ss</u>urvival times of <u>different</u> groups <u>characterized by theirof</u> Gal-1 expression in <u>the</u> nucleus, cytoplasm, and stroma <u>have beenwere</u> compared (<u>Figure 2Fig. 2</u>). Cases with Gal-1 expression in the cytoplasm showed significantly reduced overall survival compared to cases without any Gal-1 expression in the cytoplasm (p = 0.029) Moreover, cases displaying high Gal-1 expression in the stroma showed <u>a</u>-significantly <u>reduced-poorer</u> outcomes <u>compared to</u> <u>casesthan those</u> with low Gal-1 expression in the stroma (p = 0.045). <u>A Ccomparison of</u>

<u>cases ng</u> negative <u>versus and</u> positive <u>cases offor</u> Gal-1 expression in the nucleus did not show <u>reveal</u> any differences <u>with regardin terms of to</u> overall survival. However, based on <u>considering a</u>-multivariate analysis, only Gal-1 stromal staining <u>would</u>-serves as an independent prognostic factor (<u>Table 2Table 2</u>).

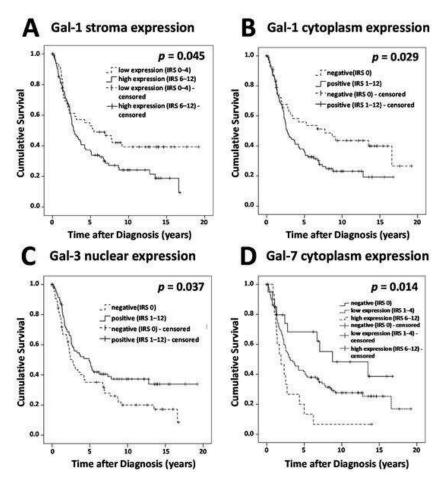


Figure 2. Survival times were plotted as Kaplan-Meier graphs. Percentage of living patients (vertical axis) was plotted <u>in dependence of against</u> time (horizontal axis). Patients without an observed event (death) who exited the study before the observation period ended have been censored, as indicated - Censoring has been marked in the graphs. Survival times of different groups of stratified by Ggalectin expression have been are compared. <u>Galectin expression was</u>

determined in the cytoplasm, nucleus, and stroma using Remmele immunoreactive (IR)-scores. (A) Cases displaying high Gal-1 expression in the stroma showed-a significantly reduced outcome-survival compared to cases with low Gal-1 expression in the stroma. (B) (A) Cases with Gal-1 expression in the cytoplasm showed significantly reduced overall survival compared to cases without any-Gal-1 expression in the cytoplasm.; (C) (B) Cases without Gal-3 expression in the nucleusi showed significantly reduced overall survival compared to cases with nuclear Gal-3 expression.; (D) (C) Cases with high Gal-7 expression showed a significantly reduced overall survival, when compared to cases with low expression of Gal-7.; (D) Galectin expression was determined in cytoplasm, nucleus, and stroma using Remmele (IR) scores.

Table 2. Multivariate analysis of prognostic factors for overall survival in ovarian cancer.-

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-	Confederation (b)	IIP End (h)	95%			
Covariate	Coefficient (b _i)	HR Exp (b _i)	Lower	Upper	p-Value	
Histology (serous vs. other)	0.211	1.235	0.658	2.317	0.511	
Grade (G1 vs. G2, G3)	0.942	2.565	1.290	5.100	0.007	
FIGO (I, II vs. III, IV)	1.140	3.126	1.537	6.357	0.002	
Patients' age (≤60 vs. >60 years)	0.312	1.367	0.861	2.169	0.185	
Gal-1 stroma (low vs. high)	0.571	1.770	1.044	2.999	0.034	
Gal-1 cytoplasm (neg. vs. pos.)	-0.187	0.830	0.423	1.626	0.586	
Gal-3 nucleus (neg. vs. pos.)	-0.265	0.767	0.480	1.227	0.269	
Gal-7 cytoplasm (neg. vs. pos.)	0.636	1.889	1.160	3.077	0.011	

HR = hazard ratio; CI = confidence interval

2.2. Presence of Gal-3 in <u>the Nn</u>ucle<u>usi</u> <u>i</u>Is <u>a</u>A <u>p</u>Positive <u>p</u>Prognostic<u>indicator</u> in <u>o</u> Θ varian <u>c</u> Θ cancer

Gal-3_positive nuclei were observed in 83 (55%) out of 151 cases, while 96 cases (63.6%) showed cytoplasmic Gal-3 staining and 85 cases (56.3%) presented with Gal-3_positive peritumoural stromae (Figure 1Fig. 1). Median IR scores for Gal-3 in the nucleusi, cytoplasm, and stroma were 1, 2, and 1, respectively. Gal-3 staining showed-was correlatedions with clinical and pathological data-variables (Table 3 Table 3). Gal-3 expression in the stroma and nucleus was differed amongnt for several different histological subtypes (p = 0.008 and p = 0.013, respectively). Gal-3 stromal staining was stronger in the serous and clear cell subtypes but weaker in the endometrioid and mucinous subtypes, while nuclear Gal-3 staining was stronger in the serous, clear cell, and mucinous subtypes but weaker in the endometrioid subtype. Tumours rated as pT1 presented with significantly stronger nuclear Gal-3 staining than those rated pT2 or higher staged cases (p = 0.042). We observed a-correlations of between Gal-3 staining in the nucleus and cytoplasm with patients⁴ age (p = 0.022 and p = 0.013, respectively), with observing higher IR scores for patients younger than 60-years. For

<u>In</u> our study panel, Gal-3 overexpression in the cytoplasm was not correlated with poorer outcomes of <u>in</u> ovarian cancer patients. <u>AlsoSimilarly</u>, Gal-3 staining in the peritumo<u>u</u>ral stroma <u>could not servewas not observed to be as</u> a prognostic factor. <u>HoweverIn contrast</u>, nuclear Gal-3 expression could serve as a positive prognostic factor (<u>Figure 2Fig. 2</u>). Cases without Gal-3 expression in <u>the</u> nucleus<u>i</u> showed significantly reduced overall survival compared to cases with nuclear Gal-3 expression (p = 0.034). According to the results of a multivariate analysis, <u>however</u>, nuclear Gal-3 staining <u>could not serve aswas not</u> an independent prognostic factor, probably due to its strong correlations with patient <u>s</u>² age, tumo<u>u</u>r stage, and histology (<u>Table 2Table 2</u>).

 Table 3. Correlations between Gal-3 staining correlated with and clinical and pathological data factors.

Clinical and Pathological Variables Histology	Gal-3 Expression Cytoplasm		p	Gal-3 Expression Stroma		p	Gal-3 Expression Nucleus		p
	neg.	pos.		neg.	pos.		neg.	pos.	
Serous	37	69	NS	42	64	0.008	44	62	0.013
Clear cell	3	.9		42	10		3		0.0000
Endometrioid		9		13				9 5 7	
Mucinous	12 3	9		13 9	83		16 5	7	
Tumor Stage									
pT1	12	27	NS	21	18	NS	12	27	0.042
pT2+	43	68		44	67		55	56	
Lymph node									
pN0/pNX	39	62	NS	47	54	NS	48	53	NS
pN1	16	62 34		19	31		20	30	
Distant Metastasis									
pM0/pMX	53	92	NS	64	81	NS	65	80	NS
pM1	2	4		64 2	4		65 3	3	
Grading									
Gi	9	28	NS	16	21	NS	13	24	NS
G2+	40	62		44	58		51	51	
FIGO									
1/11	13	30	NS	21	22	NS	15	28	NS
III/IV	41	62		43	60		51	52	
Age									
≤60 years	22	57	0.022	33	46	NS	28	51	0.013
>60 years	33	39		33	39		40	32	

TNM staging was accomplished performed according to actual the standards of the UICC; pT1 = tumour stage 1; pT2+ = tumour stage 2 or higher; pN0 = lymph node stage 0; pNX = lymph node stage not evaluated; pN1 = lymphnode stage 1; pM0 = distant metastasis stage 0; pMX = distant metastasis not evaluated; pM1 = distant metastasis stage 1; G1 = grade 1; G2+ = grade 2 or higher; NS = Not significant (p > 0.05).

2.3. Gal-7 <u>eExpression Llevels pPredicts Shortened Oo</u>verall <u>sSurvival in oOvarian cCancer</u> Staining for Gale<u>ein-</u>7 was mainly observed in the cytoplasm; only <u>a</u> few individual cases showed nuclear staining (Figure 1Fig. 1). Cytoplasmic Gal-7 staining was present in 129 (86.6%) out of 149 specimens, with a median IR score of 3. In total, 20 cases <u>presented-were</u> negative for Gal-7, while 114 cases showed low and 15 cases showed high expression of Gal-7. Gal-7 expression appeared to show differ <u>amongences for several different</u> histological subtypes (p = 0.026). The strongest Gal-7 staining was found in <u>the</u> serous subtype, and the weakest was in the endometrioid subtype (Table 4Table 4). No other correlations of between Gal-7 <u>staining and with pathological data was-were</u> found. Survival times of Gal-7_-negative cases and <u>cases those displaying awith</u> high Gal-7 expression were compared to <u>cases those</u> with low Gal-7 expression (Figure 2Fig. 2). We observed a significantly reduced overall survival for cases with high Gal-7 expression and <u>a betterimproved</u> survival for Gal-7_ negative cases_, when-compared to <u>that of</u> cases with low expression of Gal-7 (p = 0.014). AlsoIn addition, according to the results of <u>a</u>-multivariate analysis, <u>higher</u>-Gal-7 expression can be confirmed as an independent prognostic factor for overall survival in ovarian cancer (Table 2Table 2).

Clinical and Pathological Variables	Gal-7 E	p		
	neg.	low	high	
Histology	~			
Serous	10	83	12	0.026
Clear cell	0	10	2	
Endometrioid	7 3	13	2 0 1	
Mucinous	3	8	1	
Tumor Stage				
pT1	4	29	5	NS
pT2+	15	85	10	
Lymph node				
pN0/pNX	15	75	8	NS
pN1	5	39	8 7	
Distant Metastasis				
pM0/pMX	19	110	14	NS
pM1	1	4	1	
Grading				
G1	6	25	3	NS
G2+	12	80	3 11	
FIGO				
I/II	8	29	4	NS
III/IV	11	81	11	
Age				
≤ 60 years	12	59	8	NS
>60 years	8	55	8	

 Table 4. Correlations between Gal-7 staining correlated with and clinical and pathological data factors.

TNM staging was accomplished performed according to actual the standards of the UICC; pT1 = tumour stage 1; pT2+ = tumour stage 2 or higher; pN0 = lymph node stage 0; pNX = lymph node stage not evaluated; pN1 = lymphnode stage 1; pM0 = distant metastasis stage 0; pMX = distant metastasis not evaluated; pM1 = distant metastasis stage 1; G1 = grade 1; G2+ = grade 2 or higher; NS = Not significant (p > 0.05).

2.4. Correlations among galectin expression patternsAnalysis

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<u>Results of the Aanalysis of the correlations among galectin expression patternsanalysis is</u> are shown in <u>Table 5</u>. For Gal-1 staining, we observed positive correlations <u>between</u> among staining <u>results</u> in the cytoplasm, nucleus, and stroma. <u>AlsoSimilarly</u>, the staining <u>results results</u> of Gal-3 in <u>the cytoplasm</u>, nucleus, and stroma were positively correlated among with each other. Furthermore, we <u>found-observed</u> correlations between Galectin-1 and -3 staining in <u>the</u> nucleus, cytoplasm, and stroma. Gal-7 staining <u>showed-was</u> positively correlate<u>dions</u> with Gal-1 <u>staining</u> in <u>the</u> cytoplasm and nucleus and all types of Gal-3 staining.

Staining	Gal-1 Cytoplasm	Gal-1 Stroma	Gal-1 Nucleus	Gal-3 Cytoplasm	Gal-3 Stroma	Gal-3 Nucleus	Gal-7 Cytoplasn
Gal-1 cytoplasm		0.5.45.55.5	OVER 198	and the second second	510.00.007	N. N. S. L	10 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
cc	1.000	0.382	0.748	0.356	0.263	0.282	0.272
p		< 0.001	< 0.001	< 0.001	0.001	< 0.001	0.001
n	150	150	150	149	149	149	146
Gal-1 stroma							
cc	0.382	1.000	0.231	0.123	0.280	-0.006	-0.040
Р	< 0.001	and the second	0.004	0.135	0.001	0.937	0.633
n	150	150	150	149	149	149	146
Gal-1 nucleus							
cc	0.748	0.231	1.000	0.302	0.315	0.329	0.249
p	< 0.001	0.004	0.000	< 0.001	< 0.001	< 0.001	0.002
n	150	150	150	149	149	149	146
Gal-3 cytoplasm							
cc	0.356	0.123	0.302	1.000	0.293	0.839	0.276
p	< 0.001	0.135	< 0.001	2010	< 0.001	< 0.001	0.001
11	149	149	149	151	151	151	146
Gal-3 stroma							
cc	0.263	0.280	0.315	0.293	1.000	0.267	0.231
p	0.001	0.001	< 0.001	< 0.001		0.001	0.005
n	149	149	149	151	151	151	146
Gal-3 nucleus							
cc	0.282	-0.006	0.329	0.839	0.267	1.000	0.335
p	< 0.001	0.937	< 0.001	< 0.001	0.001	35	< 0.001
n	149	149	149	151	151	151	146
Gal-7 cytoplasm							
cc	0.272	-0.040	0.249	0.276	0.231	0.335	1.000
p	0.001	0.633	0.002	0.001	0.005	< 0.001	1.00
n	146	146	146	146	146	146	149

Table 5. Correlation analysis of galectin expression patterns.

<u>Correlations among</u> IR scores of Gal-1, -3, and -7 staining in different compartments were <u>correlated assessed</u> with <u>each other</u> using Spearman's correlation analysis. cc = correlation coefficient, p = two-tailed significance, n = number of patients.

3. Discussion

In this study, we assessed the prognostic value of Gal-1, -3, and -7 expression on overall survival in ovarian cancer patients. According to our data, Gal-1 staining in the cytoplasm and stroma predicts poor share a negative prognostic impact on overall survival in ovarian cancer. In accordanceConsistent with this, in vitro experiments have showned that the overexpression of Galectin-1 significantly increases migrationve and invasion behaviours inve behavior of ovarian cancer cells¹⁹ [19]. Furthermore, Gal-1 knockdown experiments in ovarian cancer cells displayed result in a reductions in cell growth, migration, and invasion. Possible mechanisms for this include the interaction of Gal-1 interaction-with H-Ras and to activateion of the Raf/extracellular signal-regulated kinase (ERK) pathway, as well as the downregulateion of matrix metalloproteinase-9 (MMP-9) and c-Jun-could have been explored as possible mechanisms. Moreover, Gal-1 overexpression could may significantly decrease the sensitivityies of ovarian cancer cells to cisplatin, illustrating reflecting a possible explanation for the decreased-reduced survival of ovarian cancer patients with increased Gal-1 expression¹⁴ [14]. Thus, Gal-1 is represents a promising new target for ovarian cancer therapy-For this purpose, and several compounds targeting Gal-1 have been introduced²⁰ [20]. OTX008, for instance, is a new compound able to binding non-covalently to Gal-1 on the side back face, was able to inhibiting the proliferation and invasion of various cancer cells lines²¹ [21]. The Aanti-proliferative effects of OTX008 correlated with Gal-1 expression across a large panel of cell lines. Moreover, OTX008 efficiently inhibited the growth of ovarian cancer xenografts in vivo²² [22].

According to the results of a-multivariate analysis in this study, only Gal-1 stromal staining eould-serves as an independent prognostic factor for overall survival. The Aaccumulation of Gal-1 in the peritumoural stroma has been described for various other tumour entities²³⁻²⁵ [23,24,25]. Some groups tried haveto investigated the mechanisms responsible for this phenomenon. In situ hybridization experiments were able to showed that fibroblast-seells, adjacent to malignant cells, express [GALal-1] mRNA, illustrating suggesting a possible explanation for peritumoural Gal-1 accumulation. AlsoIn addition, it was demonstrated that ovarian cancer cells produce Gal-1 and release it into the medium. Furthermore, conditioned medium obtained from ovarian carcinoma cells is able to-induces increased elevated Ggal-1 expression in fibroblast-seells. Both-These experiments suggest that primarily the ovarian cancer cells might-may be primarily responsible for stromal Gal-1 expression²⁶ [26]. Our exploration findings regarding of the positive correlation between Gal-1 staining in the peritumoural stroma and malignant cells is consistent with this hypothesis. However, it requires further investigations to are required to explain cases of without-Gal-1 expression in the stroma but not in cancer cells, and *-but in the stroma or* vice versa.

Several groups have suggested that higher Gal-3 expression is associated with reduced progression-free survival in ovarian cancer^{17,27} [17,27]. However, in these studies, observation detection of Gal-3 expression was limited to the cytoplasm, while and the prognostic value of nuclear Gal-3 staining has not been further studied. We could not confirm a negative

Commented [A20]: Focus area:

Focus on clinical implications of the study findings

Recommended actions:

I recommend that your Discussion better emphasize the contribution of this study to the prognostic potential of galectins in ovarian cancer. This is something that is lacking in the text.

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Commented [A22]: This description is a bit vague. Please be more specific if possible.

Commented [A23]: Typically, gene symbols--and not gene names--are italicized to distinguish them from proteins.

Commented [A24]: Please provide in-text citations for the studies mentioned here.

influence of cytoplasmic Gal-3 overexpression on overall survival for-in our study panel. On the contrary, nuclear Gal-3 staining served as a positive prognostic factor, although it was not independent of the influence of clinical and pathological parameters. Thus, it is Apparently, it is the nuclear and not cytoplasmic Gal-3 expression that has a major influence on patients' outcomes. In line with this, Gal-3 has been observed to play an important role in nuclear eell physiology, as it is involved in the mechanisms-processes of pre-mRNA_-splicing or and mRNA transport^{28.29} [28,29]. Furthermore, cell culture experiments using human cervix adenocarcinoma (HeLa) -cells showed-demonstrated a-delayed activation of the DNA damage repair response activation and a decrease in the G2/M cell cycle checkpoint arrest in the absence of Gal- 3^{30} [30]. A similar mechanism could be s conceivable in ovarian cancer, predisposing cells for further mutations in the absence of nuclear Gal-3. To our knowledge, reduced Gal-3 expression as an indicator of poorer prognosis has only been observed in gastric cancer so-thus far³¹[31]. In cholangiocarcinoma, Gal-3 expression was is associated with a poorly -differentiated type, while *in vitro* experiments showed significantly increased cell migration and invasion after suppression of Gal-3 expression³²[32]. However, for ovarian cancer, in vitro experiments showed have shown that knockdown of Gal-3 inhibits migration and invasion of cancer cells, while increasing apoptosis and sensitivity to carboplatin³³ increases [33]. Moreover, paclitaxel and additional treatment with a Gal-3 inhibitor treatment showed resulted in synergistic cytotoxic effects and increased apoptosis in an on-ovarian cancer cell line³⁴ [34]. Since there are disagreements Due to the discrepancies in previous research and to the fact that our data is are not neither consistent with either previous studies on progression-free survival nor with-recent results of in vitro research, further investigation on-into the prognostic role of Gal-3 in ovarian cancer is definitely required.

As recently proposed by other groups, we were able to confirm Gal-7 as <u>a</u> negative prognosticator for overall survival in ovarian cancer <u>in-according to both</u> uni- and multivariate analys<u>e</u>is. <u>Further eC</u>ell culture experiments <u>were able to provehave demonstrated</u> that Gal-7 expression is induced by a mutant form of p53. <u>AlsoIn addition</u>, <u>Ge</u>al-7 was shown to increase <u>the</u> proliferation¹⁶ [16], invasiveness, and motility of ovarian cancer cells, while <u>interacting as an</u> immunosuppressantive by killing Jurkat T_-cells and human peripheral T_cells¹⁸ [18]. <u>All in all Together</u>, these investigations confirm Gal-7 as a <u>new</u> promising <u>new</u> target for specific therapeutic <u>option-treatment of in</u> epithelial ovarian cancer.

We observed various a variety of positive correlations between among the expression patterns of Gal-1, -3, and -7. This observation, and along with the fact that gGalectins share binding affinities and have exhibit similarities in protein structure, suggests the assumption that Ggalectins might also share common functions in ovarian cancer molecular biology. However, since as these observations are required to explore the biological characteristics and functions of different gGalectins are required to determine their manner(s) in which they are similarities and or differences, t in specifically in regards to their role(s) in ovarian cancer. Commented [A25]: Focus area: Statement not well supported by findings

Recommended actions:

This statement is premature and does not correspond to the facts. Other studies obtained the opposite results both in terms of Gal-3 localization and cancer prognosis. This statement should be deleted; instead, the reason for this discrepancy between the present and earlier studies should be discussed.

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5. Conclusions

In this study, We were able to we showed that Galeetin expression of galectins and theirits impacts on overall survival of in ovarian cancer patients is are strongly dependent onf itstheir cellular localization, whether it is in the nucleus or cytoplasm of tumour cells or the peritumoural stroma. We elaborated found that Gal-1 tumour and stromal staining, and Gal-7 staining in the cytoplasm serves as a negative prognostic factors for overall survival in ovarian cancer, while nuclear Gal-3 staining could may serve as a positive prognostic factor. According to the results of a-multivariate analysis, Gal-1 stromal staining and Gal-7 staining are prognostic factors that are, independent of clinical and pathological parameters.

4. Materials and Methods

4.1. Patients

Formalin-fixed, paraffin-embedded (FFPE) ovarian cancer samples from 156 female patients who underwent surgery at the Department of Obstetrics and Gynecology, Ludwig-Maximilians-s--University (LMU) of Munich, Germany between 1990 and 2002 were analyszed in this study. Women diagnosed for with benign or for borderline tumours of the ovary were excluded, and no patient had received neo-adjuvant chemotherapy. Tumour grading [(G1 (n = 38), G2 (n = 53), G3 (n = 53)], and histological characterization [(serous (n = 110), endometrioid (n = 21), clear cell (n = 12), mucinous (n = 13)) were performed by a gynecological pathologist. Tumour staging was accomplished performed using FIGO classifications [(I (n = 35), II (n = 10), III (n = 103), IV (n = 3)]). TNM classification was performed according to the UICC. Data on the extension of the primary tumour was-were available in 155 cases [(T1 (n = 40), T2 (n = 18), T3 (n = 93), T4 (n = 4)], data on lymph node involvement was were available in 95 cases [(N0 (n = 43), N1 (n = 52))], and data on the presence of distant metastasis was were available in 9 cases [(M0 (n = 3), M1 (n = 6)]. Clinical data was-were retrieved from patients' charts, and follow--up data was-were requested from the Munich Cancer Registry. Patients' age at surgery ranged between from 31 and to 88 years, with a median age of 62 ± 12 years. Mean overall survival was 3.2 ± 3.0 years, and 104 deaths were observed in total. The mean follow_-up time-period was 5.1 ± 4.8 years.

4.2. Immuno<u>histo</u>chemistry

Resected ovarian cancer tissue samples were fixed in formalin and embedded in paraffin after surgery. For histopathological investigations, sections were dewaxed in <u>x</u>Xylol for 20 <u>minutes-min</u> and immersed in 3% hydrogen peroxide (Merck, Darmstadt, Germany) to quench endogenous peroxidase. Then, slides were rehydrated in a descending series of alcohol (100%, 75%, and 50%), and cooked <u>in a pressure cooker</u> for 5 <u>minutes-min</u> in sodium citrate buffer (0.1 mol/L citric acid_/0.1 mol/L sodium citrate, pH 6.0) <u>in a pressure cooker</u> to ensure epitope retrieval. Afterwards, slides were washed in distilled water and phosphate-buffered saline (PBS), followed by a specific procedure for <u>staining_each gGalectin-staining.-In</u> particular, fFor Galectin-1 (Gal-1) staining, slides were blocked using <u>Pp</u>ower <u>Bb</u>lock **Commented [A27]:** This section was moved to before the Materials and Methods section to improve readability and flow.

This section could be further enhanced by the addition of a discussion of any limitations of the study, as well as the broader implications of your study. As *Scientific Reports* caters to a broad audience, are there other fields of research that may be impacted by your study?

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Recommended actions:

The number of patients was not justified by power analysis, and it is unclear whether the sample size was sufficient to achieve statistical significance.

There appear to be no control samples, i.e., those from cancer-free individuals.

You need to address these two points prior to submission.

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(BioGenex, San Ramon, CA, USA) for 3 min at room temperature and incubated with aAnti-Gal_ectin-1 primary antibody (goat, polyclonal; R&D Systems, Minneapolis, MN, USA) at a final concentration of 0.033 µg/mL in Ppower bBlock (BioGenex, San Ramon, CA, USA) for 16 h at 4 °C. Galectin 3 (Gal-3) staining was performed by blocking specimens with 1.5% horse serum (Vector Laboratories, Burlingame, CA, USA) for 30 min at room temperature and incubating with <u>aAnti-Galectin_3</u> primary antibody (mouse, monoclonal;, Novocastra Reagents, Leica Biosystems, Wetzlar, Germany) at a final concentration of 4.6 µg/mL in PBS for 16 h at 4 °C. For Galectin 7 (Gal-7) staining, specimens were blocked with Blocking Solution [(Reagent 1,; ZytoChem Plus HRP Polymer System (Mouse/Rabbit); Zytomed Systems GmbH, Berlin, Germany]) for 5 minutes min at room temperature. Slides were then incubated with aAnti-Gal-7 (rabbit, polyclonal; Abcam, Cambridge, UK) at a final concentration of 2.5 µg/mL in PBS for 16 h at 4 °C. Afterwards, for Gal-1 and -3 staining, slides were incubated with isotype-matcheding anti-goat/mouse -IgG secondary antibody and avidin-biotin-peroxidase complex, both for 30 min at room temperature, according to the instructions of the ABC Vectastain kit (Vector Laboratories, Burlingame, CA, USA). For Gal-7 staining, specimens were incubated in <u>Ppost-Bblock</u> reagent (Reagent 2,)(Zytomed Systems GmbH, Berlin, Germany) and HRP-Polymer (Reagent 3,)-(Zytomed Systems GmbH, Berlin, Germany) for 30 min at room temperature, according to the manufacturer's protocol for the -{ZytoChem Plus HRP Polymer System (Mouse/Rabbit) (Zytomed Systems GmbH). All slides were washed twice in PBS for 2 min after every incubation step. For visualization reaction, every-specimens wereas stained with 3.3'-diaminobenzidine chromogen (DAB; Dako, Glostrup, Denmark). The reaction was, stopped after 30 s-to-2 min with tap water, and specimens were counterstained in Mayer acidic hematoxylin, dehydrated in an ascending series of alcohol followed by xylol, and covered with Consul Mount (Thermo Shandon, Pittsburgh, PA, USA). Tissue sections that had been previously incubated with isotypematched rabbit-/mouse-/goat- IgG (Dako, Hamburg, Germany) instead of the primary antibody served as negative controls. For positive controls, tissue slides of placental tissue (Gal-1, -3) or breast cancer (Gal-7) tissues were used. Primary antibodies were chosen due to the high expected staining specificities according to the results of positive--control staining, as well as descriptions, and example pictures on the manufacturers's homepages. TheA semiquantitative -method (IR score; Remmele IR score) was performed determined by two independent observers in consensus to obtain staining results. For this purpose, the predominant staining intensity (0 =negative, 1 =low, 2 =moderate, and 3 =strong) and the percentage of stained cells (0 = 0%, 1 = 1-10%, 2 = 11-50%, 3 = 51-80%, and 4 = 81-100%stained cells) are has to be multiplied, resulting in values from 0 to 12. Staining intensity was measured in the cytoplasm and the nucleus of the cancer cells, and in the peritumoural stroma. Cut-off points for IR scores were chosen specifically for each staining with regard to the distribution pattern of IR scores in the collective sample. For Gal-1 staining in the cytoplasm and nucleus of cancer cells, <u>an IR score</u> = 0 was considered as negative and an IR score $S \ge 1$ as positive. For stromal staining, Gal-1 groups of with low expression (IR score S < 5) and

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high expression (IR <u>score</u> $\$ \ge 5$) were compared. For analysis of Gal-3 staining, negative cases with an IR <u>score</u> $\$ \ge 0$ were compared to positive cases with an IR <u>score</u> $\$ \ge 1$. Gal-7 expression was grouped as negative (IRS = 0), low ($1 \ge IRS \le 4$), and high (IRS ≥ 6).

4.3. Statistical <u>a</u>Analysis

Statistical data was obtained analyses were performed using SPSS 23.0 (\pm 23, IBM, Armonk, NY, USA)-<u>statistic software. DD</u>istributions of clinicopathological variables was were tested with Cchi-Ssquare Statisticstests. Mann-Whitney *U*-tests was were used to compare the IR scores of gGalectins between-among different clinical and pathological subgroups. Correlations between-among immuno<u>histo</u>chemical staining results were calculated using Spearman's correlation analysis. Kaplan-Meier curves and <u>Log</u>-rank tests (Mantel-Cox) were used to compare survival times between-among different groups. Data are presented with-as_the mean ± standard deviation. Values of *p* < 0.05 were considered as significant.

4.4. Ethics <u>Ss</u>tatement

All tissue samples used for this study were left-over material from the archives of <u>the</u>LMU Munich, Department <u>of</u> Gynecology and Obstetrics, <u>which Ludwig Maximilians University</u>, <u>Munich, Germany, that hadwere</u> initially <u>been</u> collected for histopathological diagnostics. All diagnostic procedures had already been fully completed at the time the histopathological investigations for the current study were performed. Patients' data <u>have beenwere</u> fully anonymized. The study was approved by the Ethics Committee of LMU Munich. All experiments were performed according to the standards set <u>forth</u> in the <u>D</u>declaration of Helsinki, <u>1975</u>.

5. Conclusions

We were able to show that Galectin expression and its impact on overall survival of ovarian eancer patients is strongly dependent of its localization, whether it is in the nucleus or eytoplasm of tumor cells or the peritumoral stroma. We elaborated that Gal-1 tumor and stroma staining, and Gal-7 staining in the cytoplasm serves as a negative prognostic factor for overall survival in ovarian cancer, while nuclear Gal-3 staining could serve as a positive prognostic factor. According to the results of a multivariate analysis, Gal-1 stroma staining and Gal-7 staining are prognostic factors, independent of elinical and pathological parameters.

Acknowledgments

This study was funded by the FöFoLe program of the Ludwig-Maximilians-University of Munich for Heiko Schulz.

Author Contributions

Udo Jeschke conceived and designed the experiments; Christina Kuhn and Simone Hofmann performed the experiments; Heiko Schulz analyzed the data and wrote the paper. **Commented [A32]:** *Scientific Reports* requires a Data Availability Statement to be included in the Methods section of submitted manuscripts (see '<u>Availability of materials and</u> <u>data</u>' section for more information). Elisa Schmoeekel and Doris Mayr revised the manuscript for important intellectual content. Sven Mahner and Udo Jeschke initiated and supervised the study. All authors read and approved the final version of the manuscript.

Conflicts of Interest

The authors declare no conflict of interest.

References

Acknowledgments

<u>This study was funded by the FöFoLe program of the Ludwig-Maximilians-University of</u> <u>Munich for Heiko Schulz.</u>

Author Contributions

<u>Udo Jeschke conceived and designed the experiments; Christina Kuhn and Simone</u> <u>Hofmann performed the experiments; Heiko Schulz analyszed the data and wrote the paper;</u> <u>Elisa Schmoeckel and Doris Mayr revised the manuscript for important intellectual content</u>; <u>Sven Mahner and Udo Jeschke initiated and supervised the study; a</u>-<u>All authors read and</u> <u>approved the final version of the manuscript.</u>

Conflicts of InterestCompeting Financial Interests

The authors declare no conflict of interest.

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